

Asymmetric Dihydroxylation of Stachysterone C: **Stereoselective Synthes**is of 24-*epi*-Abutasterone

Boon-ek Yingyongnarongkul and Apichart Suksamrarn*

Department of Chemistry, Faculty of Science, Ramkhamhaeng University, Bangkok 10240, Thailand

Received 24 November 1997; revised 7 January 1998; accepted 8 January 1998

Abstract: Stachysterone C was synthesized from 20-hydroxyecdysone (20-ECD). Sharpless asymmetric dihydroxylation of this rare ecdysteroid using osmium tetroxide and a chiral ligand afforded 24-epi-abutasterone, another rare ecdysteroid, and its C-24 epimer, abutasterone. High diastereomeric excess of the former ecdysteroid was obtained when the chiral ligands dihydroquinidine 1,4-phthalazinediyl diether and dihydroquinidine 2,5-diphenyl-4,6-pyrimidinediyl diether were employed. The two C-24 epimeric ecdysteroids exhibited similar moulting hormone activity in the Musca bioassay. However, they were significantly less active than 20-ECD. © 1998 Elsevier Science Ltd. All rights reserved.

24-epi-Abutasterone (1) is a rare ecdysteroid isolated recently in a small quantity (3 mg) from a complex mixture of the methanolic extract of 2.5 kg of *Vitex canescens* root bark.¹ The structure of 1 was deduced mainly from spectroscopic (¹H and ¹³C NMR) spectral comparisons with its C-24 epimer, abutasterone (2). The ecdysteroid 2 was found in a number of plant species.²⁴ In continuation of our study on structure-activity relationships of ecdysteroids with different oxygenation at the side chain, we would like to see whether introduction of an extra hydroxyl group to the 24-position of the parent ecdysteroid, 20-hydroxyecdysone (3),

will result in any change in moulting hormone activity of 3. Also, we would like to compare the activity of ecdysteroids with a 24S-hydroxyl group (e.g., 2) with those with the 24R-hydroxyl group (e.g., 1). The rationale behind this study was initiated by our discovery of pinnatasterone (4)⁵ and canescensterone (5)⁶ from the stem barks of V. pinnata and V. canescens, respectively. Compound 4 exhibited very low moulting hormone activity, whereas compound 5 showed very high activity in the Musca domestica assay. Though it was obvious that the pyrrole 2-carboxylate moiety was responsible for such high activity of compound 5, it was logical to study biological activity related to stereochemical arrangement of the C-24 hydroxyl group. It was therefore advantageous to have enough of compound 1 for biological activity testing. In this paper we describe a concise, stereoselective synthesis of 1 using the ecdysteroid 3 as the starting material.

RESULTS AND DISCUSSION

Starting from the readily available ecclysteroid 3,6 the diacetonide 6 was prepared by the literature method.⁷ Treatment of the pyridine solution of 6 with mesyl chloride in the presence of DMAP furnished an inseparable 3:2 mixture of the olefin diacetonide 7 and 8, presumably through the mesylate 9 (Scheme).

The acetonide protecting groups in 7 and 8 were removed by treatment with 70% AcOH⁷ to the corresponding olefin mixture, which was separated by repeated column chromatography to stachysterone C (10) and 25,26didehydroponasterone A (11). It should be noted that compound 10 is a rare ecdysteroid isolated from Stachvurus praecox in 1970.8 The key step of the synthesis of 1 was the stereoselective dihydroxylation of 10 using OsO₄ in the presence of a suitable chiral ligand. According to the Sharpless asymmetric dihydroxylation of olefins, the cinchona alkaloid derivatives we decided to use as the chiral ligands were dihydroquinidine 4methyl-2-quinolyl ether (DHQD-MQE) and dihydroquinidine 9-phenanthryl ether (DHQD-PE).9 Before investigating the selectivity of the osmylation of the olefin 10 with these chiral ligands, we chose to examine the dihydroxylation of 10 without a chiral ligand and pyridine was used in this case, using a stoichiometric quantity of OsO₄. The reaction was performed in tert-BuOH-THF-H₂O (7:4:1) solvent system and the olefin 10 was transformed to 24-epi-abutasterone (1) and abutasterone (2) in high yield, with the products 1 and 2 in a ratio of 2:1 (Table 2, entry 1). The resulting ecdysteroids could be separated by careful column chromatography. The experiment performed in the absence of a chiral ligand, therefore, revealed that the diastereomer 1 was intrinsically favoured. We then investigated the stereoselectivity of the two chiral ligands, DHOD-MOE and DHOD-PE. The ratio of the ligand:OsO₄:olefin was 1.5:1.3:1. Both of these chiral ligands gave rise to the products 1 and 2 in a ratio of 4:1 (Table 2, entries 2 and 3). We then turned to the more recent "second generation" chiral ligands, dihydroquinidine 1,4-phthalazinediyl diether, (DHQD)2-PHAL, and dihydroquinidine 2,5-diphenyl-4,6-pyrimidinediyl diether, (DHQD)₂-PYR. Under the same conditions employed in the case of the foregoing "first generation" chiral ligands, the reaction in the presence of (DHQD)₂-PHAL was found to be 7:1 selective in favour of compound 1 (Table 2, entry 4), whereas that in the presence of (DHQD)₂-PYR was 6:1 (Table 2, entry 5). The high diastereoselectivity of both of the latter ligands therefore allowed stereoselective synthesis of the ecdysteroid 1 in high diastereomeric excess. In the latter two cases, the overall yield of 1 from the diacetonide 6 was 22 - 23 %. It should be noted that asymmetric dihydroxylation of the olefin 10 with a catalytic quantity of OsO₄ was also investigated, but the reaction proceeded very sluggishly, as has been reported in a different system. 10 It is worth mentioning that synthesis of 1 also provides indirect evidence for structural confirmation of the naturally occurring 24-epi-abutasterone. Also, the synthesized abutasterone (2), especially in the case of using the non-chiral ligand, could be used in the bioassay.

Scheme Reagents and conditions: a, CH₃COCH₃, p-TsOH; b, MsCl, pyridine, DMAP, 5 °C to ambient temp.; c, 70% AcOH, EtOH; d, OsO₄, ligand (see Table 2), t-BuOH-THF-H₂O (7:4:1)

In order to obtain further information of the osmylation of the olefin 10 to the ecdysteroids 1 and 2, the chiral ligands of the dihydroquinine (DHQ) series, which were expected to effect asymmetric dihydroxylation of the olefin 10 in favour of compound 2, the ligands DHQ-MQE, DHQ-PE, (DHQ)₂-PHAL and (DHQ)₂-PYR were selected. The dihydroxylation of 10 using DHQ-MQE and DHQ-PE gave, in both cases, the ecdysteroid 1 as the major product of a 10:7 diastereomer mixture (Table 2, entries 6 and 7). The diastereomer ratio changed to 4:1 in favour of the ecdysteroid 1 by performing the reaction both with (DHQ)₂-PHAL and (DHQ)₂-PYR (Table 2, entries 8 and 9). It was evident that intrinsic factor of the dihydroxylation of 10 became more significant in the latter two cases.

Biological activity. Both of the ecdysteroids 1 and its C-24 epimer, the ecdysteroid 2, were less active than 20-hydroxyecdysone (3) in the *Musca* bioassay. It was thus evident that introduction of an additional C-24 hydroxyl group to the parent ecdysteroid 3 resulted in a significant decrease in activity. As the moulting hormone activity of the two epimeric ecdysteroids 1 and 2 was not significantly different, this led to the conclusion that the presence of a C-24 hydroxyl group, either with the *R* or *S* configuration, resulted in a decrease in activity.

Table 1 ¹H NMR Data of Ecdysteroids

Н	1	2	7*	8*	10	11
	C ₅ D ₅ N	C ₅ D ₅ N	CDCl ₃	CDCl ₃	C_5D_5N	C_5D_5N
2	4.18 (m)	4.19 (m)	4.23 (m)	4.23 (m)	4.17 (m)	4.17 (m)
3	4.23 (br s)	4.23 (br s)	4.27 (br s)	4.27 (br s)	4.24 (br s)	4.23 (br s)
5	2.99 (dd,	3.00 (dd,	2.36 (dd,	2.36 (dd,	3.02 (dd,	3.02 (dd,
_	12.9, 3.5)	12, 3.6)	12.6, 4.7)	12.6, 4.7)	13.2, 3.5)	13.1, 3.6)
7	6.22 (d, 2.1)	6.24 (d, 2.1)	5.83 (d, <i>ca</i> 2)	5.83 (d, <i>ca</i> 2)	6.25 (d, 2.1)	6.26 (d, 2.1)
9	3.57 (m)	3.58 (m)	2.81 (m)	2.81 (m)	3.60 (m)	3.59 (m)
17	3.08 (t, 9.1)	2.99 (t, 8.2)	ca 2.27 [≠]	ca 2.27 [≠]	2.95 (t, 9.3)	2.93 (t, 9.1)
22	4.51 (dd,	4.08 (dd,	3.70 (dd,	3.66 (dd	3.89 (dd,	3.84 (br d,
	$9.6, 2.3)^a$	9.7, 1.8) ^b	8.2, 4.8)	9.7, 2.7)	9.7, 1.5)	10.9)
24	4.37 (dd,	4.24 (dd,	5.18 (m)		5.55 (br t, 7)	
	9, 2.7) ^a	10.1, 1.5) ^b				
26	-	-	-	4.72, 4.75	-	4.75, 4.79
				(each br s)		(each br s)
18-Me	1.22 (s)	1.19 (s)	0.79 (s)	0.79 (s)	1.21 (s)	1.22 (s)
19-Me	1.06 (s)	1.06 (s)	0.98 (s)	0.98 (s)	1.07 (s)	1.07 (s)
21-Me	1.63 (s)	1.59 (s)	$1.17 (s)^{c}$	1.15 (s) ^c	1.59 (s) ^f	$1.57 (s)^g$
26-Me	1.46 (s)	1.45 (s)	1.64 (s)	-	1.59 (s)	-
27-Me	1.47 (s)	1.50 (s)	$1.71 (s)^{d}$	1.74 (s) ^d	1.63 (s) ^f	1.67 (s) ^g
COMO	-	-	1.33 ^e , 1.33,	1.32 ^e , 1.33,	-	-
$C(Me)_2$			1.41, 1.49	1.41, 1.49		
			(each s)	(each s)		

Assigned from a mixture of compounds 7 and 8.

Table 2 Dihydroxylation of the olefin 10

Entry	Ligand	Ratio of Products 1:2	% Yield
1	pyridine	2:1ª	88
2	DHQD-MQE	4:1ª	89
3	DHQD-PE	4:1ª	88
4	(DHQD) ₂ -PHAL	7:1 ^{a,b}	84
5	(DHQD) ₂ -PYR	6:1 ^{a,b}	82
6	DHQ-MQE	10:7ª	80
7	DHQ-PE	10:7ª	80
8	(DHQ) ₂ -PHAL	4:1ª	76
9	(DHQ) ₂ -PYR	4:1ª	76

^aDetermined by ¹H NMR spectral analysis.

a-gAssignments may be reversed for signals with the same superscript.

[≠]Obscured signal.

^bDetermined by HPLC analysis.

EXPERIMENTAL

General experimental details have been described previously.¹¹

Reaction of 20-hydroxyecdysone 2,3:20,22-diacetonide (6) with mesyl chloride

Compound 6⁷ (1.5565 g, 2.779 mmol) was dissolved in pyridine (5 ml) and the solution stirred at 0-5 °C for 10 min. Mesyl chloride (1 ml, 12.867 mmol) and 4-dimethylaminopyridine (DMAP, 50 mg) were added and the reaction mixture was left to stir at 5 °C for 20 min, then slowly allowed to warm up to ambient temperature during the period of 4 h. Water was added to the reaction mixture and the solution extracted with CHCl₃ (3x35 ml). The combined CHCl₃ layer was washed with water, dried over anhydrous Na₂SO₄ and evaporated to dryness. The crude mixture was subjected to column chromatography using CHCl₃-MeOH as eluting solvent, with an increasing amount of the more polar component. Fractions eluted by CHCl₃-MeOH (99:1) afforded a 3:2 mixture of stachysterone C 2,3:20,22-diacetonide (7) and 25,26-didehydroponasterone A 2,3:20,22-diacetonide (8) (1.3160 g, 87%).

7+8: IR: v_{max} 3474, 2976, 1658, 1445, 1373, 1243, 1215, 1057, 884, 756 cm⁻¹; ¹H NMR data of compounds **7** and **8** are given in Table 1; EIMS: m/z (% rel. intensity) 542 [M][†] (2), 527 (9), 509 (12), 484 (8), 467 (7), 466 (11), 403 (34), 385 (13), 327 (7), 183 (16), 140 (9), 125 (100), 107 (35). Anal. Calcd for $C_{33}H_{50}O_6\cdot H_2O$: C, 70.71; H, 9.28. Found: C, 70.93; H, 9.39.

Acetonide deprotection of 7 and 8

To a solution of the acetonides 7 and 8 (147 mg, 0.271 mmol) in MeOH (1 ml) was added 70% AcOH (5 ml) and the mixture was stirred at ambient temperature for 4 days. The reaction mixture was poured into water and the solution extracted with *n*-BuOH (3x30 ml). The combined organic layer was washed with water, and evaporated by co-distillation with water under reduced pressure. The crude mixture which contained stachysterone C (10) and 25,26-didehydroponasterone A (11) in a ratio of 3:2 (from ¹H NMR) was chromatographed using CHCl₃-MeOH with increasing concentration of MeOH to afford pure compound 10, a mixture of compounds 10 and 11, and pure compound 11. The compounds 10 and 11 mixture was chromatographed to yield more of pure compounds 10 and 11 together with a mixture of 10 and 11, the latter of which was subjected to another column chromatography. This resulted in the separation of a total of 68 mg (54%) of compound 10, 28 mg of compound 11 and a mixture (7 mg) consisting mainly of the compound 11 and some of the compound 10.

10: Amorphous, mp 225-227 °C (from MeOH-CHCl₃); IR: v_{max} 3414, 2926, 1647, 1445, 1382, 1116, 1056, 875 cm⁻¹; ¹H NMR data is given in Table 1; FABMS (+ve): m/z (% rel. intensity) 463 [M+H]⁺ (54), 445 (29), 427 (6). Anal. Calcd for $C_{27}H_{42}O_{6}$ ·3/2 $H_{2}O$: C, 66.25; H, 9.20. Found: C, 66.21; H, 8.84.

11: Needles, mp 263-265 °C (from MeOH-CHCl₃); IR: ν_{max} 3347, 2938, 1641, 1443, 1378, 1355, 1316, 1262, 1161, 1122, 1061, 997, 949, 891, 874 cm⁻¹; ¹H NMR data is given in Table 1; FABMS (+ve): m/z 463.3057 [M+H]⁺. $C_{27}H_{42}O_6$ requires 463.3059.

Dihydroxylation of stachysterone C (10) with OsO₄ and pyridine

A solution of OsO₄ (250 mg) in THF (5 ml) was prepared and a portion (288 μl, 0.056 mmol) was added to a solution of pyridine (0.8 ml) in *tert*-BuOH-THF-H₂O (7:4:1, 4 ml). Parts of the remaining OsO₄ solution were used in subsequent asymmetric dihydroxylation of the olefin 10. After 10 min stirring, a solution of the olefin 10 (20 mg, 0.043 mmol) in *tert*-BuOH-THF-H₂O (7:4:1, 1.6 ml) was then added and stirring continued for 10 min. A 1% solution of NaHSO₃ (8 ml) was added and stirring continued for 30 min. Saturated brine (80 ml) was added; the mixture was repeatedly extracted with *n*-BuOH until no products were detected in the

aqueous phase and the combined organic phase was evaporated by co-distillation with water. The residue, the ¹H NMR spectrum of which indicated a 2:1 mixture of 24-epi-abutasterone (1) and abutasterone (2), was subjected to column chromatography, using CHCl₃-MeOH as eluting solvents, to give compound 2 (7 mg) and compound 1 (12 mg). TLC and spectroscopic (¹H NMR and IR) comparisons of 1 with 24-epi-abutasterone isolated from *V. canescens* root bark¹ revealed the identity of the compounds. Compound 1 crystallized as needles from MeOH-EtOAc, mp 257-259 °C; FABMS (+ve): m/z 497.3118 [M+H] ⁺. C₂₇H₄₅O₈ requires 497.3114.

2: Needles, mp 258-260 °C from MeOH-EtOAc (lit.² 257-259 °C); IR: v_{max} 3376, 2944, 1639, 1461, 1382, 1183, 1090, 1072, 922, 874 cm⁻¹; ¹H NMR spectral data of 2 (Table 1) were consistent with the reported abutasterone.⁴ FABMS (+ve): m/z (% rel. intensity) 497 [M+H]⁺ (7), 479 (3), 461 (3), 443 (3).

Asymmetric dihydroxylation of 10 with OsO₄ and chiral ligands

General procedure. To a solution of 0.016 mmol of the chiral ligand in tert-BuOH-THF-H₂O (7:4:1, 1.8 ml) was added the previously prepared THF solution of OsO₄ (72 μl, 0.014 mmol) and the mixture stirred for 10 min. A solution of the olefin 10 (5 mg, 0.011 mmol) in tert-BuOH-THF-H₂O (7:4:1, 0.4 ml) was then added and stirring continued for 10 min. The ratio of the ligand, OsO₄ and olefin was 1.5:1.3:1. A 1% solution of NaHSO₃ (2 ml) was added and stirring continued for another 30 min. Saturated brine (20 ml) was then added and the mixture repeatedly extracted with *n*-BuOH until no products were detected in the aqueous phase. The combined organic phase was evaporated and the residue was chromatographed to separate a mixture of compounds 1 and 2 from the ligand. The ratio of the two ecdysteroids was determined by ¹H NMR spectral analysis and, in the case of the dihydroxylation using (DHQD)₂-PHAL and (DHQD)₂-PYR where the product ratio could not accurately be determined by the ¹H NMR method, HPLC analysis (column: Spherisorb ODS2, 5 μm, 250x4.6 mm; mobile phase: MeOH-H₂O (1:3); flow rate: 1 ml min⁻¹; detector: 254 nm). The results are shown in Table 2, entries 2-9.

Biological activity testing. The *Musca* bioassay was performed by the method referred to previously.⁵ **Acknowledgements.** This work was supported by the Thailand Research Fund. We are grateful to Mrs Wanna Sririnnuth and Mr Nitirat Chimnoi for recording the NMR and mass spectra, respectively.

REFERENCES

- 1. Suksamrarn, A.; Promrangsan, N.; Chitkul, B.; Homvisasevongsa, S.; Sirikate, A. *Phytochemistry* **1997**, 45, 1149-1152.
- 2. Pinheiro, M. L. B.; Filho, W. W.; da Rocha, A. I.; Porter, B.; Wenkert, E. *Phytochemistry* 1983, 22, 2320-2321.
- 3. Zhang, M; Stout, M. J.; Kubo, I. Phytochemistry 1992, 31, 247-250.
- 4. Coll, J.; Reixach, N.; Sanchez-Baeza, F.; Casas, J.; Camps, F. Tetrahedron 1994, 50, 7247-7252.
- 5. Suksamrarn, A.; Sommechai, C. Phytochemistry 1993, 32, 303-306.
- 6. Suksamrarn, A.; Sommechai, C.; Charulpong, P.; Chitkul, B. Phytochemistry 1995, 38, 473-476.
- 7. Suksamrarn, A.; Pattanaprateep, P. Tetrahedron 1995, 51, 10633-10650.
- 8. Imai, S.; Murata, E.; Fujioka, S.; Koreeda, M.; Nakanishi, K. Chem. Commun. 1970, 352-353.
- 9. Kolb, H. C.; VanNieuwenhze, M. S.; Sharpless, K. B. Chem. Rev. 1994, 94, 2483-2547.
- 10. Tsubuki, M.; Takada, H.; Katoh, T.; Miki, S.; Honda, T. Tetrahedron 1996, 52, 14515-14532.
- 11. Suksamrarn, A.; Yingyongnarongkul, B. Tetrahedron 1996, 52, 12623-12630.